

Measurement Error Does Not Explain the Persistence of a Body Mass Index Association with Endometrial Cancer after Adjustment for Endogenous Hormones

Nancy Potischman, Mitchell H. Gail, Rebecca Troisi, Sholom Wacholder, Robert N. Hoover

Identified risk factors for endometrial cancer are accepted as operating through estrogen exposure. In a recent analysis, the effect of risk factors such as body mass index (BMI) was not explained by circulating estrogen concentrations. In the present analysis, we correct for measurement error associated with obtaining only one blood sample per subject. Applying

regression calibration ideas, we found that error correction of log estrone had little impact on estimates of the BMI effect, suggesting that hormone measurement error does not account for the residual importance of BMI. The biologic mechanism for the increased risk associated with BMI remains to be explained. (*Epidemiology* 1999;10:76-79)

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We recently published results on risk related to endogenous hormone levels in a case-control study of endometrial cancer,¹ testing the hypothesis that epidemiologic risk factors for endometrial cancer operate through endogenous hormonal mechanisms, namely circulating estrogens. Support for this hypothesis comes from studies of unopposed estrogen therapy.^{2,3} Another risk factor, obesity, is also thought to operate through estrogen mechanisms,⁴ since estrogens are produced in adipose tissue.^{5,6} These estrogens are also more bioavailable because of lower levels of sex-hormone binding globulin (SHBG) in obese compared with non-obese women.^{7,8} Other risk factors, including history of oral contraceptive use, parity, and age at menopause, also are thought to operate through circulating hormones.^{9,10} To our knowledge, our study is the first large-scale epidemiologic endeavor to collect both questionnaire and serologic data, and to evaluate the relations among risk factors.

Subjects and Methods

Our case-control study¹ involved newly diagnosed cases from seven hospitals and controls matched on age, race, and area of residence. Community controls under age 65 were obtained through random-digit dialing procedures;

those age 65 or older were derived from the Health Care Financing Administration. Women referred to these hospitals for benign gynecological conditions formed an additional control group. Results were not different for the two control groups, which we combined for these analyses. Subjects were interviewed and then measured for a variety of anthropometric indices including height and weight, from which we calculated the body mass index (BMI kg/m²).¹¹ We excluded women who reported use of exogenous estrogen within 6 months, resulting in 208 cases and 209 controls for analysis. Assays were conducted by Nichols Institute (San Juan Capistrano, CA).

Results

Presuming that endogenous estrogen levels explain the effect of risk factors such as BMI in our study, we expected that the odds ratios (ORs) for endogenous estrogens would be of substantially greater magnitude than those for the risk factors. We also anticipated that effect estimates for "surrogate" risk factors would be eliminated or substantially diminished after adjustment for an exposure variable that mediated the effect. Neither of these expectations was met in these data. For example, adjusted for other factors besides hormones, the OR for highest to lowest BMI category was 3.8 (95% confidence interval (CI) = 2.2-6.4), whereas the estimates for the highest vs lowest estrogen quartiles ranged from 3.0 for total estradiol to 3.8 for estrone without adjustment for BMI. With adjustment for BMI, the effect estimates were reduced to 1.3 and 2.2 for estradiol and estrone, respectively. In contrast, the BMI odds ratio was essentially unchanged by adjustment for any of the hormone measures. The odds ratio for BMI was barely affected even when adjusted for SHBG (OR_{BMI} = 3.1,

From the Epidemiology and Biostatistics Program, Division of Cancer Epidemiology and Genetics, National Cancer Institute, Bethesda, MD.

Address correspondence to: Nancy Potischman, Nutritional Epidemiology Branch, Division of Cancer Epidemiology and Genetics, National Cancer Institute, Executive Plaza North, Suite 430, Bethesda, MD 20892-7366.

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95% CI = 1.8–5.4), despite strong associations of SHBG with both BMI and endometrial cancer risk in these data. The effect estimates for other established risk factors (for example parity, smoking) were similarly unaffected after adjustment for endogenous hormones.

Because the hormone levels used in these adjustments were based on a single sample from each woman, we assessed whether errors in estimates of estrogen levels could account for their minimal impact on the risk estimate for BMI. We studied estrone because it is the predominant circulating estrogen in postmenopausal women. To examine the effects of errors in estrone measurement, we considered the logistic model

$$\log \{p/(1-p)\} = \mu + \beta_1 X_1 + \beta_2 X_2 + \beta_3 X_3, \quad (1)$$

where p is the risk of endometrial cancer, X_1 is the natural logarithm of true recent (assume within 3 years) estrone level (pg/mL) for a given woman, X_2 is her BMI, and X_3 is her age in years. Both X_2 and X_3 are assumed to be measured without error. Because we have only one value of estrone, which fluctuates over time for a given woman, we do not measure X_1 but rather $Z_1 = X_1 + \epsilon$, where ϵ is assumed independent of X_1 . The quantities X_1 and ϵ are assumed normally distributed with respective means μ and 0 and variances σ_1 and σ^2 . The estimated coefficients, which we obtain by including Z_1 instead of the true value X_1 in Eq 1 are $\beta_1^* = 0.44$, $\beta_2^* = 0.081$ and $\beta_3^* = 0.025$. Applying regression calibration ideas,¹² we obtain corrected estimates that account for the measurement error in Z_1 (Appendix). These corrected estimates are $\beta_1 = 0.62$, $\beta_2 = 0.075$, $\beta_3 = 0.026$. We see that correction for error in Z_1 "deattenuates" the odds ratio associated with \ln (estrone) considerably, but has only a minor impact in decreasing the apparent effect of BMI from $\beta_2^* = 0.081$ to $\beta_2 = 0.075$. We conclude that measurement error in estrone does not account for the residual importance of BMI after adjustment for \ln (estrone).

Discussion

There are several alternative explanations for the persistence of BMI as an important factor after adjustment for estrone. First, long-term postmenopausal estrone levels may be the causal factor, and current BMI may better reflect long-term estrone exposure than do current postmenopausal estrone levels. Nevertheless, the observation that excess endometrial cancer risk from exogenous estrogen diminishes within 2–5 years of cessation¹³ suggests that recent exposure is important. A second possibility is that BMI is a marker for other factors and that both estrone and these factors contribute independently to risk. One alternative pathway for a BMI effect was thought to be through insulin resistance,¹⁴ which should be a marker for a variety of metabolic aberrations.¹⁵ In these data, however, there was no main effect of c-peptide, a marker of insulin secretion, after adjustment for BMI. In addition, the odds ratio for BMI was unchanged after adjustment for c-peptide.

TABLE 1. Crude Odds Ratios by Tertile of Estrone (E1) and Quartile of BMI

BMI (kg/m ²)	Estrone (pg/mL)		
	<26	26–40	≥41
<23.0	1.00* (15)	1.49 (12)	0.78 (5)
23.0–26.0	0.87 (8)	1.07 (12)	0.93 (6)
26.1–30.0	0.73 (7)	0.70 (9)	1.87 (11)
>30.0	0.27 (1)	6.43 (31)	4.80 (90)

Numbers in parentheses indicate number of cases.

* Reference group.

Another possibility is that factors represented by BMI not only act as independent risk factors, as in Eq 1, but also interact with estrogens to elevate risk. We calculated crude odds ratios for tertiles of estrone and quartiles of BMI (Table 1). The data were suggestive of an interaction.

Two assumptions deserve mention. The estimate of intraclass correlation was based on annual measurements over the most recent 2–3 years.¹⁶ It is possible, although we believe unlikely, that errors about a woman's long-term mean are positively correlated over yearly intervals. In this case, a lower intraclass correlation might be found for measurements taken at longer time intervals, such as decades. Nevertheless, even if the intraclass correlation is 0.66, which is the lower confidence limit in the data of Hankinson *et al*,¹⁶ the value of the BMI coefficient is only reduced to 0.072 from 0.081 (an 11% reduction) (Appendix). For the BMI coefficient to be reduced by 50%, the intraclass correlation would have to be 0.3–0.4, which seems implausible. Second, we assumed a classical error model wherein the errors in the estrone measurement were independent of the true estrone value.¹⁷ Perhaps this assumption does not hold precisely, but we are unaware of data to suggest such a violation of the classical model.

Our findings, together with our earlier finding of increased risk associated with high androstenedione concentrations, suggest a need to investigate alternative explanations for the risk associated with BMI and for the etiology of endometrial cancer.

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Appendix: Calculation of Bias in the Estimate of the BMI Effect from Measurement Error in log(Estrone)

Let X_1 be the true natural logarithm of estrone, X_2 be true BMI, and X_3 be the age for a given woman. We assume that the log odds of disease is $\mu + \beta_1 X_1 + \beta_2 X_2 + \beta_3 X_3$ as in Eq 1 in the text. Suppose that BMI and age are measured without error but that a single measurement of log (estrone), Z_1 , satisfies $Z_1 = X_1 + \epsilon$, where ϵ represents a measurement error, independent of X_1, X_2, X_3 , with mean zero and variance σ^2 . The variance, σ^2 , represents both secular variation in log (estrone) within each woman and laboratory error. Letting $\Sigma = \{\sigma_{ij}\} = \{\text{cov}(X_i, X_j)\}$, assuming that X_1, X_2, X_3 and ϵ are jointly normal, and setting $Z_2 = X_2$, and $Z_3 = X_3$, we calculate the expected values

without error. From Eq A2, we find that β_1^* is attenuated by a factor $\sigma_{11.2}/(\sigma_{11.2} + \sigma^2)$, because both $\sigma_{11.2}$ and σ^2 are non-negative. Of greater interest for our problem is that the coefficient, β_2^* , of Z_2 (the BMI measurement) is biased by $\beta_1 \{\sigma^2 \sigma_{11.2}/\sigma^2 + \sigma_{11.2}\} \sigma^{12}$. Because σ^{12} is usually negative, β_2^* will usually overestimate the true value β_2 .

To estimate the bias in β_2^* , we calculated the sample covariance matrix of Z_1, Z_2, Z_3 from the sample of 209 postmenopausal women studied by Potischman *et al* (1) as $\hat{\sigma}^2 + \hat{\sigma}_{11} = 0.27435$, $\sigma_{12} = 1.14499$, $\sigma_{13} = -0.16992$, $\sigma_{22} = 37.38777$, $\sigma_{23} = -4.63604$, and $\sigma_{33} = 46.79062$. Other elements are given by symmetry. Under a nested random effects model, the variance of a Z_1 measurement is $\sigma_c^2 + \sigma_{11} + \sigma_b^2 + \sigma^2/r$, where σ_c^2 is the variance of a random batch effect (the women were analyzed in 14

$$E \begin{pmatrix} X_1 \\ X_2 \\ X_3 \end{pmatrix} \begin{pmatrix} Z_1 \\ Z_2 \\ Z_3 \end{pmatrix} = \begin{pmatrix} Z_1 - \{\sigma^2 \sigma^{11}(1 + \sigma^2 \sigma^{11})\} \{\sigma^{11}(Z_1 - \mu_1) + \sigma^{12}(Z_2 - \mu_2) + \sigma^{13}(Z_3 - \mu_3)\} \\ Z_2 \\ Z_3 \end{pmatrix} \quad (\text{A1})$$

where $\{\sigma^{ij}\}$ are elements in the inverse of the matrix Σ . In particular $\sigma^{11} = (\sigma_{11} - \Sigma_{12} \Sigma_{22}^{-1} \Sigma_{21})^{-1} = \sigma_{11.2}^{-1}$, where $\Sigma_{12} = (\sigma_{12}, \sigma_{13})$ and Σ_{22} is the covariance matrix of (X_2, X_3) . Replacing expected values of X_1, X_2, X_3 given Z_1, Z_2, Z_3 for X_1, X_2, X_3 in Eq 1, as in the regression calibration technique developed by Rosner, *et al* (12), we find that the coefficients β_1^*, β_2^* , and β_3^* of Z_1, Z_2 , and Z_3 are given by

$$\beta_1^* = \beta_1 / (1 + \sigma^2 \sigma^{11}) = \beta_1 \{\sigma_{11.2} / (\sigma_{11.2} + \sigma^2)\}, \quad (\text{A2})$$

$$\beta_2^* = \beta_2 - \beta_1 \{\sigma^2 \sigma_{11.2} / (\sigma^2 + \sigma_{11.2})\} \sigma^{12}, \quad (\text{A3})$$

and

$$\beta_3^* = \beta_3 - \beta_1 \{\sigma^2 \sigma_{11.2} / (\sigma^2 + \sigma_{11.2})\} \sigma^{13}. \quad (\text{A4})$$

These formulas extend in an obvious way for any number of independent variables X_2, X_3, \dots, X_k measured

batches), σ_{11} is the variance of the true long term log_e (estrone) levels among these women, σ_b^2 is the variance from year to year variation in a given woman (see Ref 16), and σ_c^2/r is the variance from laboratory replication error on a given sample with r replicates. An analysis variance test for $\sigma_c^2 = 0$ gave $F_{13,195} = 0.08$ with $p = 0.66$, and a moment estimator of σ_c^2 was -0.0039 . We concluded that batch effects can be ignored in this analysis, and we set $\sigma_c^2 = 0$. As mentioned above, the variance of Z_1 was 0.27435, which we equate to $\hat{\sigma}_{11} + \hat{\sigma}_b^2 + \hat{\sigma}_c^2/r$. Hankinson *et al* (16) studied 80 postmenopausal women who each provided one blood sample in 1989-1990 and two subsequent blood samples spaced approximately at yearly intervals. Assuming no batch effect, we find that the intraclass correlation estimated by Hankinson *et al* is $\text{ICC} = \sigma_{11} / (\sigma_{11} + \sigma_b^2 + \sigma_c^2/r)$. Because Hankinson *et al* used the same laboratory

(Corning-Nichols Institute) as Potischman *et al*, it is reasonable to suppose $\sigma_c^2 = 0$ in the study by Hankinson *et al* and that σ_r^2/r was the same in both studies. From the value $\text{ICC} = 0.74$ with 95% confidence interval (0.66, 0.83) given by Hankinson *et al*, we estimate $\hat{\sigma}_{11} = 0.74 \times 0.27435 = 0.20302$ and $\hat{\sigma}^2 = \hat{\sigma}_b^2 + \hat{\sigma}_r^2/r = 0.27435 - 0.20302 = 0.07133$. From $\{\hat{\sigma}_{ij}\}$ we calculate $\hat{\sigma}_{11.2} = 0.16794$.

Estimates $\hat{\beta}_1^*$, $\hat{\beta}_2^*$, and $\hat{\beta}_3^*$ were obtained from Eq 1 with $Z_1, Z_2 = X_2$, and $Z_3 = X_3$ replacing X_1, X_2 and X_3 , as 0.4374, 0.08092, and 0.02547, respectively. Using the

previous estimates of σ^2 , $\{\sigma_{ij}\}$, and $\sigma_{11.2}$ in Eq A2, A3, and A4, we invert these equations successively to obtain error-corrected estimates $\hat{\beta}_1 = 0.6232$, $\hat{\beta}_2 = 0.7524$ and $\hat{\beta}_3 = 0.02558$. Note that $\hat{\beta}_1$ exceeds $\hat{\beta}_1^*$ by 42%. More important for our purposes, $\hat{\beta}_2$ is less than $\hat{\beta}_2^*$ by an amount 0.0057, equivalent to a 7% decrease. As a sensitivity analysis, we assumed $\text{ICC} = 0.66$, which is the lower confidence limit of Hankinson *et al*, and obtained $\hat{\beta} = 0.07238$, which is 11% less than $\hat{\beta}_2^*$. Thus, measurement error in Z_1 does not explain the persistent effect of BMI after adjustment for \log_e (estrone) in model (1).